

Figure 3. (A) Orthogonal cross-sections of a T2-weighted image depicting an infarcted canine myocardium. (B) Toroidal glyph field color-coded with TV (%). (C) Toroidal field color-coded with the fiber inclination angle. Notice the rearrangement of fiber orientation within the infarcted region into a more vertical pattern as shown by lighter blue and red toroids.

A new toroid-based representation of diffusion tensor fields is proposed to improve depiction of myofiber structure and defined an innovative diffusivity map, the toroidal volume. Results indicate that the toroidal representation improve myocardial structure characterization compared to conventional representations.

1082. FREE-BREATHING SINGLE-SHOT DENSE MYOCARDIAL STRAIN IMAGING USING DEFORMABLE REGISTRATION

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Introduction

Free-breathing scans are often desirable in patients who find breath-holding difficult. We present a new approach for free-breathing myocardial strain imaging with displacement-encoding (DENSE)(1). It acquires images with a single-shot sequence and removes respiratory motion using deformable registration.

Methods All volunteers (n=4) were scanned on a 1.5T Siemens Avanto scanner. A multi-slice, single-shot DENSE sequence (Fig.1) with true-FISP readout (Fig.2)(2, 3) and FOV-selective excitation(4) was used to acquire 3 short-axis slices in a heartbeat. Ramped flip-angles through the readout train equalized the echo amplitudes. Displacement-encoding and image acquisition were placed at end-systole and late-diastole respectively, in order to capture the accumulated wall strain of systolic contraction. Imaging parameters were true-FISP echo spacing of 2.5 ms, matrix size of 128×40, FOV of 512×120mm with restricted excitation (equivalent to a matrix of 128×96 with 3/4 phase-encode FOV), slice thickness of 6-8mm, and ECG triggering every 2 RR intervals. A data set contained 270 images of 3(slices)×3(encoding directions)×30(repetitions). The encoding directions were oblique and combined in-plane

encoding moments of 4.0 mm/radian with a through-slice (Z) moment of 1.0 mm/radian. This Z moment was sufficient to suppress the unwanted DC and conjugate-echo signal in the single-shot sequence(4, 5). Each direction was acquired in a separate heartbeat. The total scan time was 3 - 4 minutes. Image registration used a deformable algorithm(6). The registered images produced 90 strain maps= 3(slices)×30(repetitions). These were averaged over the repetitions to produce average strain maps for the slices. A correlation threshold was used to remove occasional images of poor registration quality due to ECG mistrigger. To evaluate strain noise reduction by this approach, we calculated the spatial variance of the pixel-wise strain values in the LV wall for three processing configurations: 1. single repetitions without averaging; 2. the average strain maps; 3. average strain map of respiratory gated (RG) data by selecting 1/3 of the repetitions with the highest diaphragm positions (end expiration).

Results A typical circumferential strain map from a single repetition and the corresponding average strain map of all repetitions show marked difference in the noise level (Fig.3). The spatial variance of strain decreased from 0.127 to 0.063 from single repetition to the average of the RG group, and further down to 0.054 upon registration and averaging of all repetitions. The remaining variance may account for real variation around the LV.

Conclusion and Discussion

Deformable registration of single-shot DENSE images permits averaging over all of the respiratory cycle for noise reduction. Although the resolution of 4.0mm is low, the pixel-wise measurement obviates the need for ROI analysis.

References

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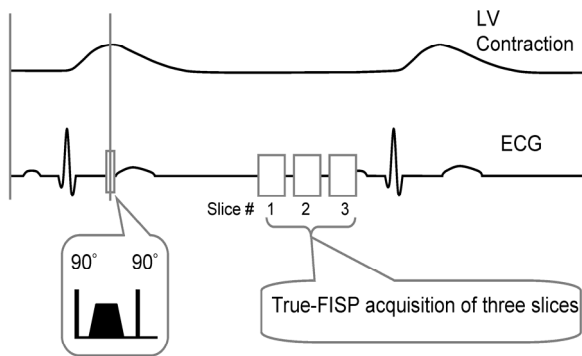


Figure 1 Time line of left ventricle contraction, ECG and the sequence.

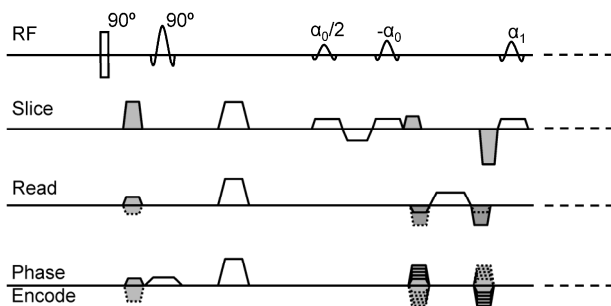


Figure 2 True-FISP DENSE sequence.

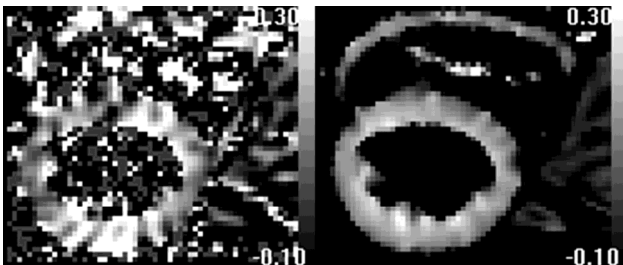


Figure 3 Left: Strain map from a single measurement; Right: strain map from the average of all registered images.

Elastic registration of single-shot DENSE myocardial strain maps allows the patient to breathe freely and permits averaging over multiple repetitions to reduce the noise level of the strain measurement. The effectiveness of this technique is shown in volunteers.

1083. REPRODUCIBILITY OF RIGHT ATRIAL VOLUME AND EJECTION FRACTION IN HEALTHY SUBJECTS AND PATIENTS WITH RIGHT HEART FAILURE USING THE STANDARD SHORT AXIS AND AREA-LENGTH METHOD

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Introduction: Reproducibility of right atrial volume and ejection fraction in healthy subjects and patients with right heart failure using the standard short axis and area-length method Burkhard Sievers, Marvin Addo, Frank Breuckmann, Joerg Barkhausen, Raimund Erbel

Background: Measurements of atrial volumes and ejection fraction (EF) are superior to diameters for both accurate determination of the atrial size and follow up studies. Changes of right atrial volume and EF might have a prognostic impact in patients with right heart failure. We therefore sought to evaluate the reproducibility of right atrial volumes and EF in healthy subjects and patients using the standard short axis method (SA) and the rapid area-length method (ALM). **Methods:** Right atrial volumes (maximum and minimum) and EF were measured in 10 healthy subjects and 10 patients with right heart failure using SA and ALM. Images were acquired with a steady state free precession gradient-echo sequence on a clinical 1.5 Tesla magnetic resonance scanner (Siemens, Erlangen, Germany). For SA, volumes were determined by the sum of the outlined areas. EF was calculated as follows: $EF = (EDV-ESV)/EDV \times 100$. For ALM, the right atrial area and length were measured from the horizontal long axis view. Minimum and maximum volumes were calculated as follows: $8 \times (\text{Area})^2 / 3\pi \times \text{Length}$, $EF (\%) = (\text{Maximum volume} - \text{Minimum volume}) / \text{Maximum volume} \times 100$. All patients were examined twice (scan 1 and 2). Both scans were performed at the same day.

Results: For SA, maximum volumes, minimum volumes and EF for healthy subjects were $95.4 \pm 19.9 \text{ mL}$, $47.9 \pm 8.9 \text{ mL}$, $49.0 \pm 8.1\%$ in scan 1 and $95.8 \pm 17.5 \text{ mL}$, $49.5 \pm 11.2 \text{ mL}$, $48.1 \pm 8.8\%$ in scan 2 ($p \geq 0.285$). SA-volumes and EF for patients in scan 1 and 2 were $145.2 \pm 28.2 \text{ mL}$, $106.9 \pm 25.9 \text{ mL}$, $26.5 \pm 9.7\%$ and $146.3 \pm 26.3 \text{ mL}$, $109.9 \pm 23.9 \text{ mL}$, $24.9 \pm 9.8\%$, respectively ($p \geq 0.139$). SA-interstudy variability was $-0.3 \pm 7.9 \text{ mL}$, $-1.6 \pm 4.9 \text{ mL}$ and $0.9 \pm 3.5\%$ for healthy subjects (Figure 1A) and $-1.1 \pm 6.8 \text{ mL}$, $-3.0 \pm 5.2 \text{ mL}$ and $1.7 \pm 2.7\%$ for patients, respectively (Figure 1B). For ALM, maximum volumes, minimum volumes and EF for healthy subjects were $89.3 \pm 19.4 \text{ mL}$, $43.7 \pm 8.0 \text{ mL}$, $50.1 \pm 8.3\%$ for scan 1 and $81.7 \pm 15.8 \text{ mL}$, $38.7 \pm 7.5 \text{ mL}$, $51.8 \pm 10.3\%$ for scan 2 ($p \leq 0.114$). ALM-volumes and EF for patients in scan 1 and 2 were $139.9 \pm 28.3 \text{ mL}$, $103.7 \pm 26.9 \text{ mL}$, $26.5 \pm 10.3\%$ and